

# The Role of Dynamic Stimulation Pattern in the Analysis of Bistable Intracellular Networks

Th. Millat<sup>a</sup>, S.N. Sreenath<sup>b</sup>, R.P. Soebiyanto<sup>b</sup>, J. Avva<sup>b</sup>, K.-H. Cho<sup>c</sup>, and O. Wolkenhauer<sup>a</sup>

Bistable intracellular networks play an important role in the functioning of living cells. Besides the well-established steady-state properties, some important characteristics of bistable systems arise from their dynamics. We demonstrate that:

- a supercritical stimulus amplitude is not sufficient to move the system from the lower to the higher branch for either a step or a pulse input
- a switching surface can be identified for the system as a function of the initial condition, input pulse amplitude and duration (a supercritical signal)
- a minimal signal power is necessary to change the steady state of a bistable system
- we investigate and characterize the role of the duration of the stimulus

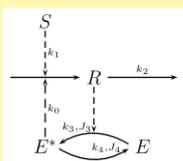
These results are relevant for the design of experiments, where it is often difficult to create a defined pattern for the stimulus.

## Model System

- Mutually-activated system [1]
- Linear system coupled with a sigmoidal system through a positive feedback loop
- Mathematical representation

$$\frac{dR}{dt} = k_0 E^*(R) + k_1 S - k_2 R$$

$$E^*(R) = G(k_3 R, k_4, J_3, J_4)$$



- Response component  $R(t)$  and external stimulus  $S(t)$
- Kinetic constants  $k_i$  and Michaelis-Menten constants  $J_i$  determine the chemical properties of the involved species
- Modified form of the enzyme  $E^*(R)$  facilitates the production of  $R$
- (De)modification of enzyme  $E$  is assumed to be at steady state which is described by the Goldbeter-Koshland function  $G(\cdot)$  [1,2]

$$G(k_3 R, k_4, J_3, J_4) = \frac{2k_3 R J_4}{\mathcal{X} + \sqrt{\mathcal{X}^2 - 4(k_4 - k_3 R)k_3 R J_4}}$$

$$\mathcal{X} = k_4 - k_3 R + k_4 J_3 + k_3 R J_4$$

- Typical sigmoidal shape  $E^*(R)$ , Fig. 1 (left)
- High nonlinear behaviour can cause multiple steady states
- Positive feedback loop regulates the production rate of  $R$  in an autocatalytic fashion

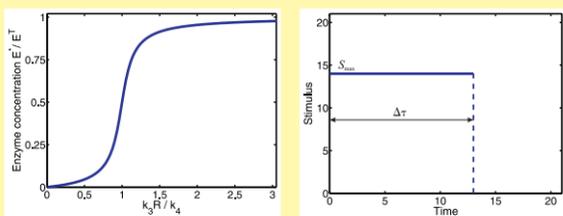


Fig. 1: Left - Concentration of the modified enzyme  $E^*$  as a function of the response component  $R$ ; Right - External stimulus with amplitude  $S_{max}$  and duration  $\Delta\tau$  which is used for the following investigations.

- Constant stimulus with amplitude  $S_{max}$  and duration  $\Delta\tau$ , Fig. 1 (right)

$$S = S_{max} [u(t) - u(t - \Delta\tau)]$$

- Step function  $u(t)$  [3]
- Pulse if  $\Delta\tau$  is finite
- Stepwise if  $\Delta\tau = \infty$

## Steady-State Properties

- Steady state with step input
- Determined by the balance equation

$$k_0 E^*(R_{SS}) + k_1 S_{max} - k_2 R_{SS} = 0$$

- Balance equation of a bistable system has at least three bio-physically realizable solutions
- Two stable branches,  $R_{SU}$  (upper) and  $R_{SL}$  (lower), are separated by an unstable one,  $R_{UN}$ , Fig. 5

$$R_{SS} \in \{R_{SU}, R_{UN}, R_{SL}\}$$

- Unstable and unphysical states define a forbidden region, Fig. 2
- Depending on the feedback strength, the network can be bistable in a reversible or irreversible manner [1] or monostable

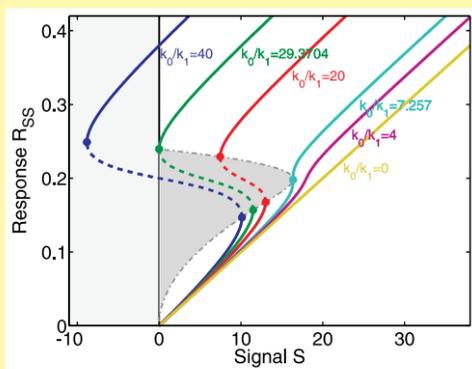


Fig. 2: Stimulus-Response curve for varying feedback strength. The system changes its behaviour from irreversible to reversible bistability and finally to monostability. The branches are separated by critical points (circles).

- Stability can be determined by the Jacobian of the system

$$J = \frac{\partial}{\partial R} \frac{dR}{dt} = k_0 \frac{\partial E^*(R)}{\partial R} - k_2$$

$$= \begin{cases} \text{negative for: } R_{SU} \text{ and } R_{SL} \text{ (stable)} \\ \text{positive for: } R_{UN} \text{ (unstable)} \end{cases}$$

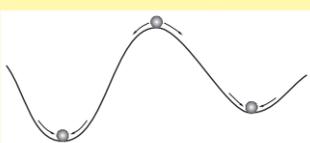


Fig. 3: Schematic representation of the stability of steady states. The system returns to the same stable steady state after a small perturbation and moves to a stable state if the unstable state is perturbed.

## References

- [1] J.J. Tyson *et al.*, *Curr. Opin. Cell Biol.* 15, 221 (2003)
- [2] A. Goldbeter *et al.*, *Proc. Natl. Acad. Sci USA* 78, 6840 (1981)
- [3] Bracewell, R., *The Fourier Transform and its Applications*, McGraw Hill (1999)
- [4] T. Millat *et al.*, *BioSystems* 92, 270 (2008)

## Dynamic Behaviour

- Transient dynamics of the irreversible bistable network for different initial states  $R_0$  and different supercritical stimulus pulses of different durations  $\Delta\tau$
- Supercritical constant stimulus  $S_{max} > S_{crit}$  always switches the system to the upper stable branch
- Systems behaviour for subcritical stimulus  $S_{max} < S_{crit}$  depends on  $R_0$ , Fig. 4

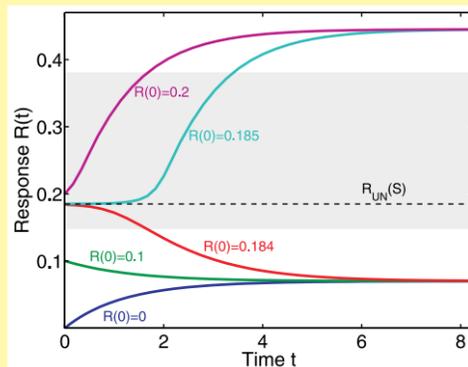


Fig. 4: Relaxation of the system for a constant subcritical stimulus  $S_{max} < S_{crit}$  for different initial states. Depending on the initial state  $R_0$ , the bistable network approaches the lower or upper branch.

- Unstable branch  $R_{UN}$  acts as separatrix for subcritical signals
- Upper branch is approached for  $R_0 > R_{UN}$ , otherwise the lower branch, Fig. 5

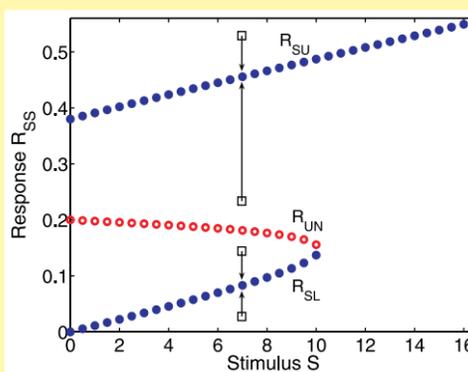


Fig. 5: Relationship between initial conditions  $R_0$  (squares), stable steady states ( $R_{SU}$ ,  $R_{SL}$ ), unstable steady state ( $R_{UN}$ ) and the corresponding critical value of step amplitude  $S_{crit}$ .

- Response to a supercritical pulse depends on the duration  $\Delta\tau$ , Fig. 6
- A critical duration  $\Delta\tau_{crit}$  is necessary in order to switch the system
- Transient response has to exceed the unstable branch before the stimulus becomes subcritical

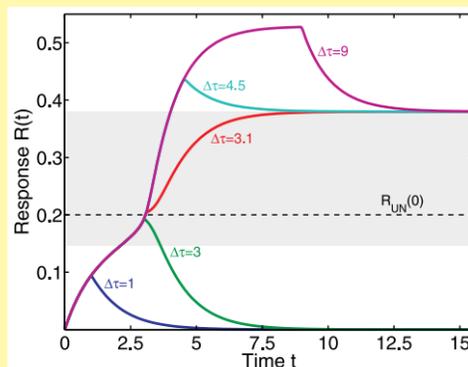


Fig. 6: Transient pulse response to supercritical stimulus  $S_{max} > S_{crit}$  as a function of the duration  $\Delta\tau$  from the origin. If  $R$  exceeds the unstable state during this time it approaches the upper branch after the pulse is switched off. Otherwise the response returns to the lower branch.

## Switching Surface

- Signal amplitude, signal duration, and initial state can be combined in a three dimensional plot, Fig. 7
- Switching surface separates the parameter combinations which switch the irreversible bistable network to the upper stable branch and which not
- Different characteristic behaviour are identified

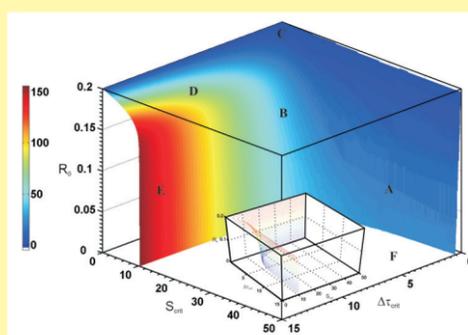


Fig. 7: Switching surface of the bistable network as response to the pulse. Regions A-F represent characteristic characteristics of the surface. Heatmap color refers to the critical power [4].

- A: Low values of  $R_0$  and  $\Delta\tau_{crit}$  require a high value of the pulse amplitude.
- B: With ever so slight increase in  $\Delta\tau_{crit}$ , and for the same low values of  $R_0$ , a much lower value of the pulse amplitude is needed for switching.
- C: At a fixed but high level of  $R_0$ , the switching is independent of critical pulse duration and only a low level of stimulus amplitude is needed.
- D: For amplitudes  $10 > S_{max} > 8$  the switching surface slopes faster along the initial condition range  $R_0 = [0.15, 0.18]$ .
- E: Between  $10.4 > S_{max} > 10.1$ , the switching surface rapidly falls indicating that the system can switch for smaller values of  $R_0$  for given  $\Delta\tau_{crit}$  and ever so small increase in  $S_{max}$ , finally having the ability to switch at  $R_0 = 0$ .
- F: This flat region indicates that for  $R_0 = 0$ , the system can switch for a wide range of an area spanned by  $S_{max} \in [10.4, 50]$  and  $\Delta\tau_{crit} \in [1, 15]$ .

## Critical Signal Duration

- The original system of differential equations cannot be solved analytically due to the high nonlinear characteristics of the Goldbeter-Koshland function  $G(\cdot)$
- Taylor expansion to approximate the enzyme concentration  $E(R)$  [4]

$$E^*(R) = \frac{J_4 - k_3 R}{1 + J_3 k_4} + \frac{1 + J_3 + J_3 J_4 k_3^2}{(1 + J_3)^3} R^2 + \mathcal{O}(R^3)$$

$$= C_1 R + C_2 R^2 + \mathcal{O}(R^3),$$

- Linear approximation with respect to response component  $R$
- Positive feedback decreases the degradation of  $R$
- Linear differential equation

$$\frac{dR}{dt} = k_1 S_{max} - (k_2 - k_0 C_1) R$$

- Analytical solution can be transformed with respect to critical signal duration [4]
- Decreased degradation rate constant  $k'_2$  and displacements between initial state  $R_0$  and stable steady state  $R_{SS}$  and unstable state  $R_{UN}$ , respectively, determines the critical duration

$$\Delta\tau_{min} = \frac{1}{k'_2} \ln \frac{R_{SS} - R_0}{R_{SS} - R_{UN}} \quad k'_2 = k_2 - k_0 C_1$$

$$R_{SS} = k_1 S_{max} / k'_2$$

- Good agreement for high stimuli, Fig. 8
- Deviations for small supercritical stimuli because of neglect of higher feedback terms in the above expansion
- Asymptote for strong stimuli  $S_{max} \gg k'_2 R_{UN} / k_1$

$$\Delta\tau_{min} \approx \frac{R_{UN} - R_0}{k_1 S_{max}}$$

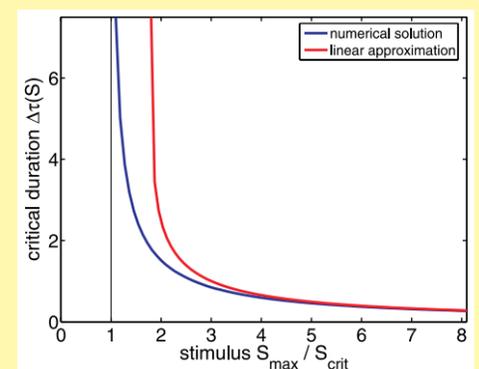


Fig. 8: Critical duration  $\Delta\tau_{crit}$  as a function of the stimulus amplitude  $S_{max}$ . Comparison of the results of a numerical calculation with the linear approximation.

## Critical Signal Power

- Signal power (dosage) sums over the whole applied external stimulus

$$P = \int_0^\infty S(t) dt$$

- $P_{crit}$  measures the dosage which is required to switch the system from a lower to a upper stable state applying a constant stimulus  $S_{max}$

$$P_{crit} = S_{max} \cdot \Delta\tau_{crit}$$

- Linear approximation and numerical results agree for high stimuli, Fig. 9
- Minimal dosage  $P_{min}$  is required for switching the system
- For strong stimuli it follows

$$P_{min} = \frac{R_{UN} - R_0}{k_1}$$

- independent from the signal strength
- determined only by rate coefficient  $k_1$ , but independent from  $k'_2$  which describes the relaxation of the system

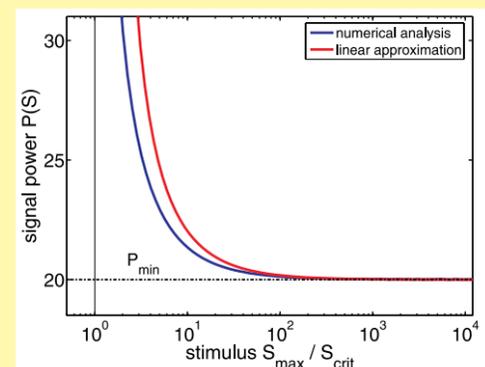


Fig. 9: Semilogarithmic plot of  $P_{crit}$  as a function of the stimulus amplitude  $S_{max}$  for the nonlinear system and the linear approximation. As pointed out in the text, there is a minimal signal power required to switch the system from the lower to the upper branch. Using the asymptotic behaviour for strong stimuli this limit can be evaluated. Interestingly it is independent from the feedback strength.

## Acknowledgements

T.M. acknowledges the support by the German Federal Ministry for Education & Research (BMBF) as part of the SysMo-Network. O.W. was supported by the European Community as part of the FP6 project AMPKIN and by the German Research Foundation (DFG). S.N.S., R.P.S., and J.A. acknowledge the support by the US National Institute of Health (NIH). K.-H. Cho was supported by the Korea Science and Engineering Foundation (KOSEF), the Korea Ministry of Science & Technology, and the 21C Frontier Microbial Genomics & Application Center Program.

## Affiliation & Contact

- Thomas.Millat@uni-rostock.de  
University of Rostock  
Institute of Computer Science  
Bioinformatics & Systems Biology Group  
18051 Rostock, Germany
- Case Western Reserve University  
Complex Systems Biology Center  
Cleveland OH44106-7071, USA
- Korea Advanced Institute of Science & Technology (KAIST)  
Department of Bio and Brain Engineering & KI for the BioCentury  
Daejeon 305-701, Republic of Korea